

# Microbiology (3)

## Bacterial Cell Wall

### Transpeptidation

- final step in cell wall biosynthesis — formation of cross-links between muramic acid residues in adjacent glycan chains
- achieved mainly through action of penicillin-binding-protein (PBP) — which catalyses transglycosylation and transpeptidation rxns. (formation of glycosidic & peptide bonds of PG)
  - ↳ FtsI in *E. coli* (PBP3)
- controlled cutting of cell wall reqd — done by autolysins which can also result in cell death

### SEDS (Shape, Elongation, Division, Sporulation)

- widespread family of bacterial cell wall polymerases
- work together as a subcomplex with the Transpeptidase PBPs.

→ RodA — elongasome (responsible for elongation of the cell)

- moves in a directed and circumferential manner around long axis of cell, polymerising PG

- FtsW - a homologous enzyme, part of divisome complex, is important for cell division, working with PBPs as well
- SpoVE - expressed in sporulating cells, resp. for spore cortex P<sub>6</sub> synthesis.  
(bac. memb. bound)
- FtsZ - bacterial homolog of tubulin, in E. coli, initiating cell division.

## Key functions of Peptidoglycan

- determines cellular shape that is reproduced from generation to generation.
- serves as attachment sites for virulence factors & adhesins
- aids bacteria in undergoing morphological transformations in response to diff. stress related factors
- in many gram +ve bacteria, PG move outward allowing cellular expansion.

## Bacillus subtilis

- has glycan strands up to 5µm, longer than cell  
- part of PG architecture allowing cell growth and division
- inner surface of cell wall has a regular macrostructure with ≈50 nm-wide peptidoglycan cables  
↳ run across the short axis of cell
- cross striations along each cable are also present
- fundamental cabling architecture is also maintained during

septum development as part of cell division.

How does peptidoglycan get its dynamic, stress-bearing structure?

- glycan is oriented parallel to the plasma memb.
- helical cabling arrangement — of cross striations

Two sets of machinery

To allow cell elongation

To the septum for cell division

- Atomic force microscopy represents cell cylinder cabling architecture — as a result of localisation of PG biosynthetic machinery
  - ↳ MreC reqd. as part of cylinder elongation biosynthetic apparatus reqd. for long glycan strands and cabling architecture.
- It is proposed that during biosynthesis, small no. of glycan strands are polymerised and cross-linked to form a PG rope
- rope is then coiled into a helix with a width of  $\approx 50$  nm to form the inner surface cable structures
- nascent helix (cable) is inserted into the cell wall by cross-links b/w two existing cables & the overlying cable interface cleaved by autolysins

- the turgor pressure of the cell causes nascent cable helix made of peptidoglycan ropes to flatten, resulting in cross-striations
- str. may get stabilised by intra/inter glycan cross-links
- helical features brought into relief during ATG hydrolysis
- accommodates very long glycan strands.

## Teichoic Acid

- have a backbone of polyol-phosphate usually with sugars and/or D-alanine as substituents
  - polyol is usually ribitol (C<sub>5</sub>) or glycerol (C<sub>3</sub>)
  - present in gram +ve bacteria, but similar polymers also occur in gram-negative bacteria (capsule or part of LPS)
  - Wall teichoic acid (WTA) and memb. lipoteichoic acid (LTA)
    - ↓
    - covalently linked to P<sub>0</sub>
    - ↓
    - anchored to plasma memb.
  - made by tag genes & tar genes
    - ↓
    - polyglycerol phosphate
    - ↪ polyribitol phosphate
- ↪ 60% of cell wall

## Function of teichoic acid:

- regulation of cell morphology and division
- scaffolding roles
- roles in cell elongation and division
- Bacteria lacking WTAs grow slower than wild types
  - clump in solution
- WTAs form a dense network of negative charge on gram- $\oplus$ ve cell surfaces — bind cationic groups (mono- & divalent-cations)  $\downarrow$

cation homeostasis - reserve for ions close to cell surfaces, important for enzymatic activity

$\Downarrow$   
WTA production is upregulated when metal  $\downarrow$

- serve as scaffolds for wide range of molecules  
(e.g., autolysins, in cell growth & div)  
 $\hookrightarrow$  grow slower in absence of WTAs

- D-alanylation reduces repulsion b/w WTA acid chains by adding  $\oplus$ vely charged amines

also increases susceptibility to antibiotics, in absence

promotes better adhesion to host tissue & confers resistance to lytic enzymes provided by host

- WTA: biosynthetic enzymes associate with protein complexes involved in elongation; and LTA with division

↓

- \* WTA in rod shaped B. subtilis → round cells  
↳ increases temperature sensitivity

- XWTA → slower growth
  - clumping
  - non-uniform thickening of PG
  - increased cell size
  - defects in septal positioning & number
  - aberrant shape
  - increased temperature & antibiotic sensitivity
  - decreased adhesion to surfaces
  - unable to grow in high salt media

### Wall teichoic acids in Streptococcus aureus

- PG is highly cross-linked - synthesised by Penicillin-binding protein (PBP-4)
- WTA → attached to PG: act as temporal and spatial regulator of PG metabolism, by localising PBP4
- In S. aureus lacking TagO, PBP4 no longer accumulates at the specifically at division septum, but is dispersed through cell membrane, as WTAs are not produced

Important: WTAs themselves are used for localisation of PBP4 and not TagO

WTA acid synthesis starts — signalling for further processing of PG — recruitment of PBP4

Hypothesis:

Bactoprenol carries both WTA and PGN;

When ↓ WTA, more bactoprenol for PGN ⇒ more delocalised substrate deloc.

[RULED OUT EXPERIMENT]

## SUMMARY

### Gram-positive cell walls

- Thick peptidoglycan
- Teichoic acids
- In acid-fast cells, contains mycolic acid

### Gram-negative cell walls

- Thin peptidoglycan
- No teichoic acids
- Outer membrane
  - LPS
  - O - polysaccharide
  - Lipid A

## Teichuronic Acid

- teichoic acids are phosphate rich, low-P substituent produced in ↓ P environments

- synthesised by tua operon — Phosphate-free acidic polymer containing GalNAc & D-glucuronic acid
- Functionally interchangeable with teichoic acids??

Indispensability of teichoic acid in context of host interaction and immune modulation

- does not contribute significantly to str. integrity & autolysin regulation.
- less anionic content.

## Micrococcus luteus

- TUA made of D-glucose & ManNAcA
- located on cell surface & covalently linked to PGR in cell wall through linker.
- synthesised by ① Glucosyltransferase  
② ManNAcA-transferase
- Teichuronic acid synthetase (TUAS) enzyme complex
  - ↓
  - 2 glycosyltransferases
  - ↳ displays hydrophobic prop. & associated with cytoplasmic memb.



## Actinoplanes spp.

characterised by

- ① aerial mycelia
- ② presence of sporangia
- ③ spherical and motile spores
- ④ variety of 2° metabolites
- ⑤ gram ⊕ve, soil, riverbed & lake sediments.

## Novel teichuronic acid in Actinoplanes lobatus

- linear str. of chain and heterogenous repeating units
  - ↳ glycopyranose residues
  - ↳ alternating residues of diaminoauronic acids - D-manno, L-gulo & D-gluco config. (6:3:1)
- not found in other gram ⊕ve bacteria

Wall Teichoic Acid	Lipoteichoic acid
◦ covalently attached to PG <sub>1</sub> layer via disaccharide phosphate residues	◦ anchored to cytoplasmic membrane through glycolipid moiety & extends into cell wall
◦ repeating units of glycerol or ribitol phosphate	◦ conserved backbone of polyglycerol phosphate

<ul style="list-style-type: none"> <li>◦ structural integrity of cell wall, cell shape, binding of cations &amp; surface protein, antibiotic resistance</li> </ul>	<ul style="list-style-type: none"> <li>◦ cell envelope integrity by inhibiting autolysin, host immune modulation (virulence factor)</li> </ul>
<ul style="list-style-type: none"> <li>◦ binds divalent cations</li> </ul>	<ul style="list-style-type: none"> <li>◦ contributes to negative charge affects cation binding</li> </ul>
<ul style="list-style-type: none"> <li>◦ forms biofilm</li> </ul>	<ul style="list-style-type: none"> <li>◦ maintains biofilm</li> </ul>
<ul style="list-style-type: none"> <li>◦ maintains localised pH for autolysin func. as well as localises autolysin to regions of division in cell wall</li> </ul>	<ul style="list-style-type: none"> <li>◦ specific inhibitor of autolysin, → resistance to lysis during stationary phase and bacterial chain formation at lower conc.</li> </ul>

## Penicillin-binding proteins (PBPs)

High Molecular Mass  
(HMM)

→ responsible for peptidoglycan polymerisation, cross-linking and insertion into pre-existing cell wall

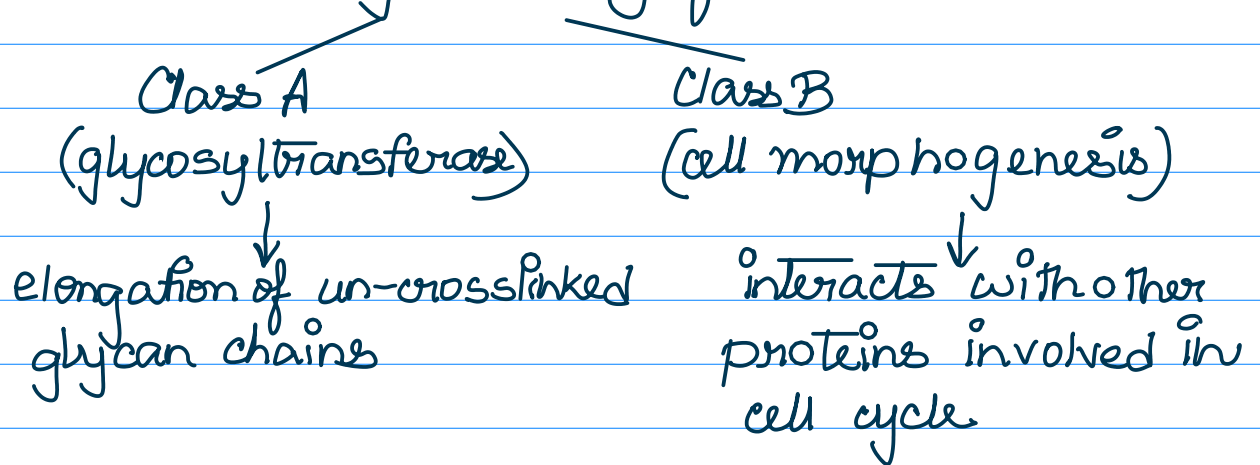
Low molecular mass (LMMs)



## Structure (HMMS)

- topology consists of a cytoplasmic tail, a transmembrane anchor, and two domains joined by  $\beta$ -linker on the outer surface of cytoplasmic memb where PG synthesis takes place

- structure & catalytic activity of N-terminal domain

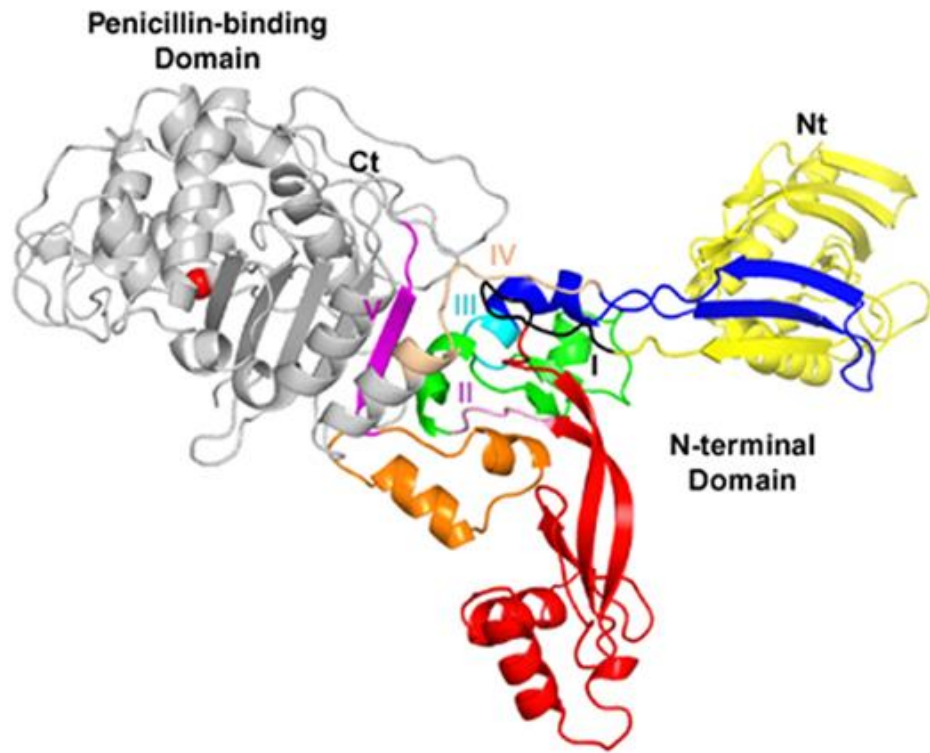


- C terminal of both classes has transpeptidase activity  
→ crosslinking b/w two adjacent glycan chains

LMM PBP → cell separation, peptidoglycan maturation or recycling

**Inside-to-outside** growth model for gram  $\oplus$ ve bacteria - new material inserted on inner face of wall, adjacent to PBPs.





Bacillus subtilis phosphate starvation

Phosphate starvation



Induction of  
Pho regulon

- induced by PhoP & PhoR
- enables cells to use limiting phosphate resources more efficiently
- genes → phoA & phoB  
(alkaline phosphatase)  
+ phoD (alkaline phosphodiesterase)

Induction of  
S<sup>B</sup>-dependent  
general stress regulon

+ pst (phosphate transport operon)

+ tua (teichuronic acid synthesis operon)

